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PLZF play as an indirect facilitator of thymic retention for the innate-like T-cells to acquire innate-like functions

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Dear editors,

Innate-like T-cells can be placed in-between the adaptive and innate immune systems. The mechanisms that determine the differentiation of innate-like T-cells is not completely understood^{1,2}. Innate-like T cells include invariant Natural Killer T-cells (iNKT), which express a $\alpha\beta$ TCR, and $\gamma\delta$ NKT cells, which express a $\gamma\delta$ TCR corresponding to V δ 6.3 and V γ 1.1. Previous study showed that Zbtb16 (PLZF) was necessary for the acquisition of an innate-like phenotype in iNKT and $\gamma\delta$ NKT cells³⁻⁵. Absence of PLZF severely impairs iNKT cell development, leading to a reduction of iNKT cell numbers. iNKT cells that develop in PLZF-deficient mice have a naive phenotype, lost the ability to co-express IL4 and IFN- γ , lost the ability to migrate to the liver and preferentially located to lymph nodes^{3,4}. Reciprocally, transgenic PLZF expression was sufficient to confer an effector phenotype to T-cells and similar migratory properties as iNKT cells⁶⁻⁸. This phenotypic conversion occurred during development and in the absence of agonist selection, indicating that PLZF expression was sufficient to alter the phenotypic characteristics of T-cells⁹. $\gamma\delta$ NKT-cells expressing V γ 1.1 and V δ 6.3/6.4 share characteristics of iNKT cells, and in PLZF-deficient mice V γ 1.1 V δ 6.3 $\gamma\delta$ T-cells were still present in reduced numbers⁹. Furthermore, our previous study demonstrated that PLZF controlled the development of fetal-derived IL-17⁺ V γ 6⁺ $\gamma\delta$ T-cells¹⁰. However, how

PLZF expression is regulated and how it exerts these functions is not clearly understood. We, therefore, decided to focus our efforts in understanding how PLZF expression provides an innate-like phenotype to iNKT cells and $\gamma\delta$ NKT cells.

At first we decided to evaluate the regulation of PLZF expression during T-cell development by PLZF-GFP reporter mice. Briefly, the reporter mice were generated by control the expression of transgene eGFP with PLZF regulatory elements in a modified bacterial artificial chromosome¹¹. We observed GFP fluorescence in iNKT cells (Fig. S1A). PLZF expression was low in early thymic progenitors (ETP), was slightly upregulated at the DN2a stage of development and turned off after T-cell specification at the DN3, DN4, and DP positive stages (Fig. S1B). We observed that contrarily to adult mice, PLZF expression was abundant in fetal thymocytes at every developmental stage (Fig. S1C). In light of the abundant expression of PLZF that we observed in the fetal thymus, we wanted to evaluate if fetal HSC may be biased to differentiate towards innate-like iNKT and $\gamma\delta$ NKT cells as compared to adult HSCs. We tested this by performing mixtures of congenic Fetal liver and Adult Bone Marrow chimeras and analyzed if iNKT and $\gamma\delta$ NKT cells were preferentially derived from fetal or adult precursors. We observed that there was no preferential bias of fetal HSCs to give rise to innate-like cells under these conditions (Fig. S1D-F).

We next tested the possibility of innate-like cells being derived from fetal precursors by performing transplants of neonatal day 1 thymus under the kidney capsule of congenic hosts. In this system, cells from the donor transplanted thymus are progressively replaced by differentiating thymocytes derived from host HSCs. Therefore,

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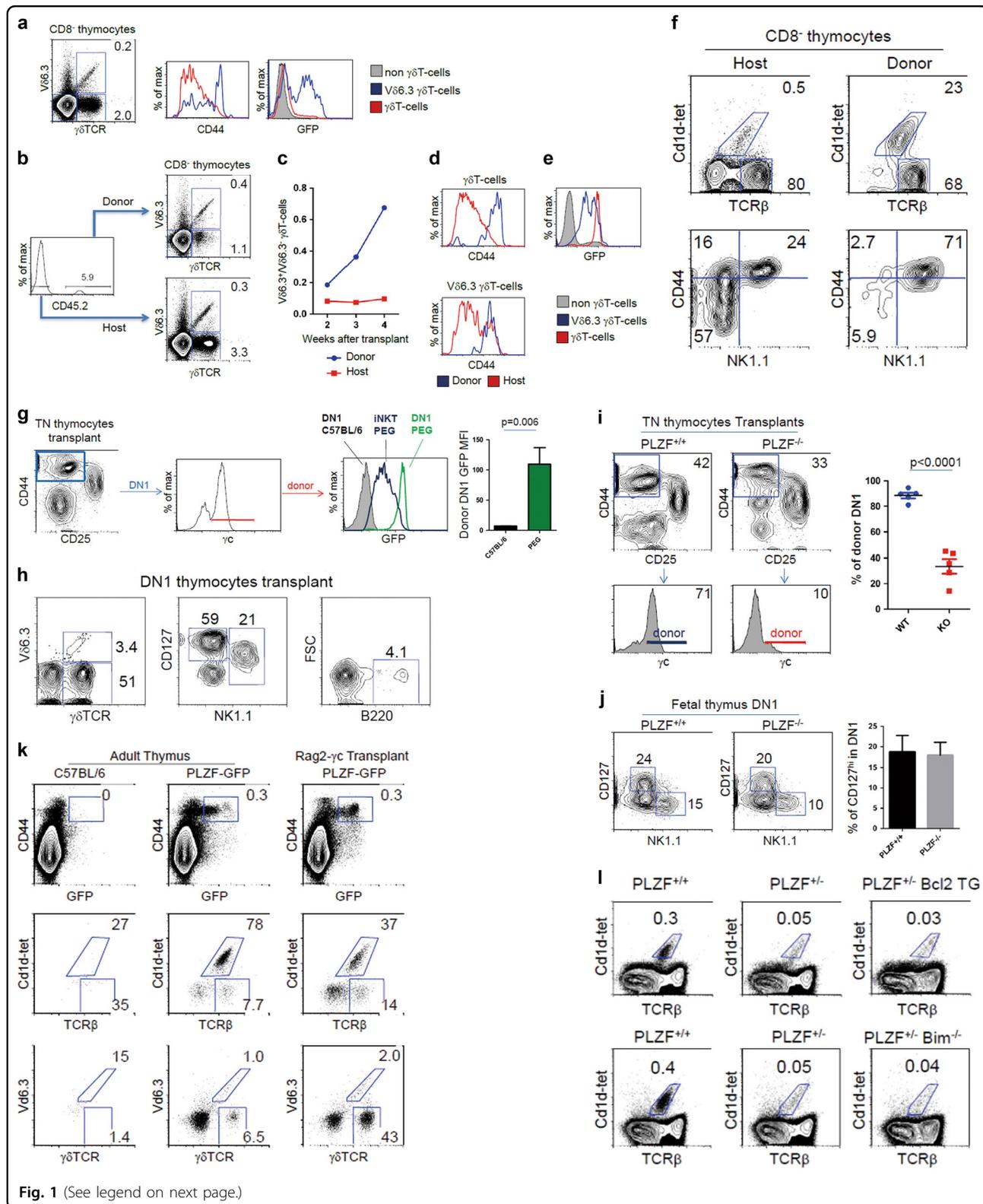


Fig. 1 (See legend on next page.)

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Fig. 1 PLZF function in thymic retention as a mechanism for the acquisition of innate-like functions. **a** Proportion of $V\delta 6.3^{+}$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells in adult PLZF-GFP thymus. CD44 and GFP levels in $V\delta 6.3^{+}$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells in adult mice. Background staining with the $V\delta 6.3$ antibody is observed in non- $\gamma\delta T$ -cells. **b** Three weeks after neonatal thymic transplants into congenic hosts. The proportion of $V\delta 6.3^{+}$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells is shown from host (CD45.2) and donor (CD45.2⁺) cells. **c** Ratio of $V\delta 6.3^{+}/V\delta 6.3^{-}$ cells derived from donor or host cells at different weeks after transplantation. **d** Analysis of CD44 levels in $V\delta 6.3^{+}$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells derived from donor or host cells. **e** Analysis of GFP levels in donor $V\delta 6.3^{+}$, $V\delta 6.3^{-}$ thymocytes and non- $\gamma\delta T$ -cells. **f** Increased retention of donor iNKT cells after thymic transplantation into congenic hosts. **g** Triple negative (CD4⁻CD8⁻TCR β) profile of transplanted thymus into Rag2- γc -deficient hosts. Gate on the DN1 (CD44⁺CD25⁻) population identify cells derived from the donor thymus (γc^{+}) and host HSC (γc). Comparison of GFP levels between PLZF-GFP (PEG) and C57BL/6 transplanted thymus gating on the TN, CD44⁺ γc^{+} donor population in relation to the GFP levels of iNKT cells in PLZF-GFP (PEG) mice. **h** Characterization of the PLZF⁺ DN1 population found in the Rag2- γc transplants according to different markers. **i** Analysis of the TN (CD4⁻CD8⁻TCR β) profile in thymic transplant of day 1 neonates from wild-type or PLZF-deficient thymus into Rag2- γc -deficient hosts. The levels of γc expression in the CD44⁺ population indicates if these cells are donor or host derived. **j** Proportion of CD127⁺ cells among DN1 thymocytes in day 15 fetal thymus from wild-type and PLZF-deficient mice. **k** FACS analysis of adult C57BL/6; adult PLZF-GFP thymus; and PLZF-GFP neonatal transplants into Rag2- γc hosts. Analysis of the proportion of iNKT (Cd1d-tet⁺) and $\alpha\beta T$ -cells, and of $\gamma\delta NKT$ ($V\delta 6^{+}$) and $\gamma\delta T$ -cells on CD44⁺GFP⁺ gated cells. **l** Protection from apoptosis in Bim-deficient or Bcl2 transgenic mice does not revert the thymic iNKT phenotype of PLZF heterozygous mice

analysis of the different T-cell subtypes in the transplant derived from donor cells is indicative of the combined ability of these cells to differentiate and to remain in the thymus. We analyzed how iNKT and $\gamma\delta NKT$ would differentiate in this system. At first, we confirmed that $V\delta 6.3^{+}$ $\gamma\delta TCR$ ($\gamma\delta NKT$) in PLZF-GFP reporter mice presented high CD44 levels and a proportion of them were GFP positive, indicative of PLZF expression (Fig. 1a). We then set-up thymic transplants of day 1 neonatal C57BL.6 thymus (CD45.2) under the kidney capsule of congenic Ly5.2 (CD45.1) mice, and analyzed the presence of $V\delta 6.3^{+}$ $\gamma\delta NKT$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells in the transplants that were derived from either donor or host cells. Three weeks after transplantation, approximately 6% of the cells in the transplanted thymus was of donor origin. Donor cells showed an increased proportion of $V\delta 6.3^{+}$ $\gamma\delta NKT$ cells among the $\gamma\delta T$ -cell population (Fig. 1b). This increase of donor $V\delta 6.3^{+}$ $\gamma\delta NKT$ cells was observed at 3 and 4 weeks after transplantation (Fig. 1c). To our surprise, high CD44 expression was not exclusive to $V\delta 6.3^{+}$ $\gamma\delta NKT$ cells and both donor $V\delta 6.3^{+}$ and $V\delta 6.3^{-}$ $\gamma\delta T$ -cells in the transplant, but not those derived from the host, had homogeneously high levels of CD44 (Fig. 1d). In correlation with high CD44 levels, donor $V\delta 6.3^{+}$ $\gamma\delta NKT$ cells as well as $V\delta 6.3^{-}$ $\gamma\delta T$ -cells expressed PLZF (Fig. 1e). Similarly to $\gamma\delta NKT$ cells, donor iNKT cells were preferentially retained in the transplants and had a mature CD44⁺NK1.1⁺ phenotype (Fig. 1f).

We were curious to interrogate if the PLZF expressing cells that we observed in the fetal thymus may remain in the adult and maintain PLZF expression if placed under non-competitive conditions in the absence of adult progenitors. To test this, we performed similar thymic transplant experiments of neonatal PLZF-GFP reporter thymus into Rag2/ γc double-deficient recipients mice. Analysis of transplants from PLZF-GFP reporter mice showed a population of DN1 thymocytes that expressed

high levels of PLZF (Fig. 1g). These CD44⁺PLZF⁺ cells showed a mature phenotype, were heterogeneous, and corresponded mostly to CD127⁺ (IL7R α ⁺), $\gamma\delta T$ -cells and NK1.1⁺ cells (Fig. 1h).

To evaluate if PLZF confers the ability to thymocytes to remain in the thymus, we set-up transplants of wild-type and PLZF-deficient neonatal thymus under the kidney capsule of Rag2- γc -deficient host mice. One month after transplantation, we observed that PLZF-deficient transplants had a severe reduction of donor CD44⁺ cells, as most of the cells found with this phenotype were derived from the Rag2/ γc hosts and were negative for the common gamma chain of the IL-2R (γc) (Fig. 1i). As fetal wild-type and PLZF-deficient thymus had a similar thymic profile, $\gamma\delta T$ -cells¹⁰ and proportion of DN1 CD127⁺ thymocytes (Fig. 1j), this led us to postulate that these CD127^{hi} cells, although present in the fetal thymus, were unable to remain in the PLZF-deficient transplants. Independent to these results, we have observed in the PLZF-GFP thymic transplants into Rag2- γc -deficient hosts that among the CD44⁺GFP⁺ populations were many $\alpha\beta T$ -cells that were not iNKT cells (Cd1d-tet⁻) and $\gamma\delta T$ -cells that were not $\gamma\delta NKT$ cells ($V\delta 6.3^{-}$) (Fig. 1k). Using this gating strategy, we were also able to detect these cells in adult PLZF-GFP thymus, although in a lower proportion (Fig. 1k).

As CD44⁺ donor thymocytes from wild-type mice in the thymic transplants did not preferentially incorporate BrDU (data not shown), these results indicated that these DN1 CD127⁺ cells were not actively dividing. Another possible mechanism that could explain the absence of these cells in PLZF-deficient transplants could be by increased apoptosis of cells in the absence of PLZF. However, we think this unlikely due to the inability of bcl2 transgenic expression or Bim deficiency, both which protect from apoptosis, to revert the deficient iNKT phenotype in PLZF heterozygous mice (Fig. 1l).

Altogether, our results suggest that PLZF play a function in the thymic retention of lymphocytes with an innate-like phenotype. As iNKT cells that express the highest levels of PLZF have not yet acquired innate-like features, our results open the possibility of PLZF in mediating thymic retention as a determinant for innate-like differentiation.

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Conflict of interest

The authors declare that they have no conflict of interest.

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