

<https://doi.org/10.1038/s41541-024-00985-y>

Filovirus vaccines as a response paradigm for emerging infectious diseases



Andrea Marzi & Heinz Feldmann

Nowadays, filovirus vaccine development may be seen as a paradigm for our response capabilities to emerging and re-emerging infectious diseases. Specifically, the West African Ebola virus disease (EVD) epidemic accelerated countermeasure licensure for several vaccine and therapeutic products. Those products have been successfully used to control EVD outbreaks in Central Africa over the past years. This positive development, however, has not yet reached beyond EVD. Therefore, it is pertinent to increase our efforts in the development of countermeasures for other human pathogenic members of the family *Filoviridae* as they continue to threaten public health in Sub-Saharan Africa. This review article summarizes the current filovirus vaccines in preclinical macaque studies and human clinical trials and discusses the most promising recent advancements.

Filoviruses are a family of enveloped, non-segmented, negative-strand RNA viruses and are mainly endemic to Africa. The family is divided into several virus genera, but outbreaks of human disease have only been caused by ebolaviruses and marburgviruses¹. Infection with the most prominent members—Ebola virus (EBOV) or Marburg virus (MARV)—causes hemorrhagic disease with case fatality rates of up to 90%¹. Outbreaks of human disease are believed to originate from zoonotic spillover events from the reservoir, likely bats, or other end host species such as nonhuman primates (NHPs), both sources of nutrition in the endemic areas. Bats are the reservoir of MARV² and have long been speculated to be the reservoir for ebolaviruses¹. EBOV and MARV recently caused disease outbreaks in areas outside of their known endemic zones which may be attributed to an expanding interface between reservoir species and the human host driven by changes in the environment, economy, public health and human behavior. Human-to-human transmission through persistently infected survivors has been suspected for a long time and recently been confirmed; however, its contribution to causing future outbreaks remains unclear but a worrying probability^{3,4}.

The 2013–2016 West African EBOV disease (EVD) epidemic accelerated countermeasure development and licensure for several products specific to EBOV in the diagnostic, vaccine, and treatment sector. As a result, two EBOV vaccines—Ervebo (based on vesicular stomatitis virus (VSV); also known as VSV-EBOV or rVSV-ZEBOV)⁵ and the prime-boost combination of Zabdeno (based on adenovirus (Ad); also known as Ad26-EBOV) and Mvabea (based on modified vaccinia Ankara (MVA); also known as MVA-BN-filo)⁶—have been approved for human use by US, European and African authorities (Table 1). In recent years, these vaccines along with the monoclonal antibody-based therapeutics Ebanga (also known as ansumivab; single monoclonal antibody)⁷ and Inmazeb (also

known as REGN-EB3; cocktail of 3 monoclonal antibodies)⁸ have been successfully used to combat EVD outbreaks in the Democratic Republic of the Congo (DRC)⁹. However, these approved vaccines and treatments are specific to EBOV with limited to no use against other ebolaviruses and MARV¹⁰. It is therefore pertinent to continue the development of countermeasures for other ebolaviruses, like Sudan virus (SUDV) and Bundi-bugyo virus (BDBV), and MARV as they continue to cause human disease outbreaks albeit with lower case fatality rates (SUDV ~50%, BDBV ~33%)¹¹ and pose a threat to public health in Africa¹².

Several vaccine platforms have been explored harnessing their advantages for rapid onset of protection and/or durability of the protective immune response. VSV-based vaccines are live-attenuated viral vectors administered as single-dose vaccines known to elicit protective immunity within 7–10 days of vaccination^{13–15}. This vaccine platform has also demonstrated limited post-exposure efficacy in NHPs for filoviruses^{16,17}. The cAd3 vaccines have also demonstrated fast-acting potential in NHPs, however, no confirmative human data are yet available¹⁸. Other platforms used to develop multi-dose approaches include protein-based subunit vaccines¹⁹ and non-replicating viral vectors including heterologous prime/boost candidates²⁰ known to elicit durable immunity.

Preclinical efficacy studies in a variety of animal models are critical for the clinical development and licensure of filovirus vaccines. While rodent models (mice, hamster and Guinea pigs) are excellent screening models for filoviruses, ultimately NHP studies are needed to establish the protective efficacy of a filovirus vaccine²¹. This fact was recently highlighted when a bivalent EBOV and SUDV vaccine achieved great immunogenicity in NHPs and humans yet failed to provide any protection against lethal SUDV challenge in the macaque disease model²².

Table 1 | Summary of species-specific filovirus vaccines with preclinical efficacy in NHP models

Vaccine	Target Species	Antigen used	Approach	Status	Product	Comments
VSV-EBOV & Ad5-EBOV	EBOV	EBOV-Makona GP	Prime-boost	Licensed in Russia; 2015	GamEvac-Combi	Largely limited to use by Russia
Ad5-EBOV	EBOV	EBOV-Makona GP		Licensed in China; 2017		Largely limited to use by China
VSV-EBOV	EBOV	EBOV-Kikwit GP	Prime	Licensed in USA, Europe, Africa; 2019	Ervebo	In use and recommended by WHO
Ad26-EBOV & MVA-BN-filo	EBOV	Ad26: EBOV-Mayinga GP MVA: EBOV-Mayinga GP SUDV-Gulu GP MARV-Musoke GP TAFV NP	Prime-boost	Licensed in Europe, Africa; 2020	Zabdeno & Mvabea	In use and recommended by WHO
cAd3-SUDV	SUDV	not disclosed		Clinical trials, phase I		Clinical development supported by BARDA
VSV-SUDV	SUDV	SUDV-Boniface GP	Prime	Preclinical		International AIDS Vaccine Initiative
ChAdOx-biEBOV	EBOV & SUDV	EBOV-Makona GP & SUDV-Boniface GP		Clinical trials, phase I		Development ceased
VSV-BDBV	BDBV	BDBV GP	Prime	Preclinical		Currently not a priority
VSV-TAFV	TAFV	TAFV GP	Prime	Preclinical		Not a priority – only a single non-lethal human case known
VSV-MARV	MARV	MARV-Angola GP	Prime	Clinical trials, phase I		Public Health Vaccines
VSV-MARV	MARV	MARV-Musoke GP	Prime	Preclinical		International AIDS Vaccine Initiative
VSV-N4CT1-MARV	MARV	MARV-Angola GP	Prime	Preclinical		Auro Vaccines
cAd3-MARV	MARV	MARV-Angola GP	Prime & prime-boost	Clinical trials, phase I		Sabin Vaccine Institute

Increased surveillance and advanced sequence technologies have resulted in the discovery of novel filoviruses allowing for virus isolation (Lloviu virus, LLOV)²³ or generation of recombinant viruses using reverse genetics systems (Bombali virus, BOMV)^{24,25}. However, little is known about the pathogenic potential of these bat-derived viruses in other species including humans complicating countermeasure development and highlighting a knowledge gap that needs to be addressed.

With the approval of EBOV vaccines and therapeutics, the focus in the filovirus countermeasure field has shifted to SUDV and MARV²⁶ as well as pan-filovirus approaches to enable a quick public health response to control filovirus disease outbreaks. This review summarizes the current state of filovirus vaccine development with a focus on preclinical macaque and human clinical trial data for vaccine candidates with trajectory towards approval.

EBOV vaccines

Globally, there are currently 4 EBOV vaccines which were approved by regulatory authorities in the wake of the 2013–2016 West African EVD epidemic. Russia was the first country to approve a heterologous prime-boost vaccine consisting of VSV-EBOV (based on the VSV glycoprotein-deleted platform, VSVΔG) and Ad5-EBOV (GamEvac-Combi) in 2015 for emergency use. Then, in 2017, China approved its Ad5-based EBOV vaccine²⁷. Finally, in 2019, the European and US authorities approved two vaccines, VSV-EBOV and the prime-boost combination Ad26-EBOV/MVA-BN-filo²⁸. Approvals for both vaccines have recently been updated to include children 12 months and older in addition to adults^{28,29}. Numerous countries in Africa and the WHO have approved the use of the latter two vaccines for broader population-based vaccination with the VSV-EBOV vaccine also authorized for use as an emergency vaccine during outbreaks^{30,31}. This vaccine is fast-acting in NHPs¹³ and humans¹⁴ and has recently been shown to indeed reduce mortality by 50% in vaccinated people during outbreaks³². EBOV vaccine development did not cease with the approvals of the above products as it remains the main cause of filovirus disease outbreaks and is included in all multivalent or pan-filovirus vaccine efforts discussed below.

SUDV vaccines

The 2022 SUDV outbreak in Uganda³³ brought this virus into the spotlight of filovirus research and particularly countermeasure development. A WHO panel ranked the available vaccines in the following order of priority: the VSV-SUDV (based on the VSVΔG platform), the chimpanzee Ad (cAd or ChAd)-based SUDV-specific vaccine cAd3-SUDV, and the ChAd-based EBOV/SUDV bivalent vaccine, ChAdOx-biEBOV¹². Clinical-grade vaccine doses of ChAdOx-biEBOV, cAd3-SUDV, and VSV-SUDV were shipped to Uganda for emergency use under a clinical trial protocol³⁴ and arrived ~80 days after the outbreak was declared¹². Despite all these efforts, the international response was too slow, and the outbreak ended before vaccinations could be implemented. Critique with the response to this outbreak has been voiced³⁵ including the need for faster vaccine deployment in preparation for future outbreaks. Of the three vaccines shipped to Uganda in 2022, there are two vaccine platforms ahead in development: cAd3-SUDV and VSV-SUDV. The ChAdOx-biEBOV vaccine did not demonstrate a protective benefit in a recent preclinical NHP study and development has ceased³⁶. However, a phase 1 clinical trial was conducted (NCT05079750) with data not yet being released. The cAd3-SUDV vaccine has been shown to protect macaques from lethal SUDV challenge³⁷ and is immunogenic with an acceptable safety profile at doses of 1×10^{10} and 1×10^{11} particle units (PU) in a recently published phase 1 clinical trial (NCT04041570)³⁸. The single dose vaccination elicited antigen-specific antibodies in 78% of clinical trial participants within 2 weeks of vaccination and were durable up to 48 weeks. Neutralizing titers and T cell responses were also detected during the trial in most participants³⁸. The VSV-SUDV vaccine protects NHPs within 4 weeks when administered by itself or in combination with other VSV-based vaccines^{39,40}. Currently, this vaccine is in clinical development. Finally, there is also an adjuvanted SUDV GP-based subunit

vaccine in development which is protective as a fresh and thermostable formulation after 3 doses in macaques^{19,41}. This vaccine is moving towards clinical development.

BDBV vaccines

BDBV is the latest of the human-pathogenic ebolaviruses. It was identified in the Bundibugyo district in Western Uganda following an outbreak of hemorrhagic disease in 2007 in humans with a case fatality of ~40%^{42,43}. In 2012, BDBV re-emerged in the DRC causing an outbreak with ~54% case fatality rate⁴⁴. Early after the initial outbreak, a macaque study showed some yet limited cross-protection against BDBV with VSV-EBOV or VSV-Tai forest virus (TAFV)⁴⁵. This study was challenging to interpret, as BDBV infection is not uniformly lethal in macaques—a contrast to other filoviruses. In 2013, a VSV-BDBV vaccine expressing the BDBV GP was developed on the VSVΔG platform and tested in macaques. A single dose of this vaccine was shown to uniformly protect macaques from BDBV infection and lethal disease⁴⁶. Even though this virus has not re-emerged, vaccine development should continue as BDBV is known to be pathogenic for humans.

TAFV vaccines

Associated with only a single, non-fatal human case of disease contracted in Côte d'Ivoire, efforts in studying TAFV have been limited since its discovery in 1994⁴⁷. However, with several filovirus outbreaks occurring in West Africa in the recent decade^{33,48}, TAFV is of differential diagnostic importance and remains a pathogenic filovirus in humans. VSV-TAFV (based on the VSVΔG platform) is the only vaccine studied for protective efficacy in macaques against TAFV challenge so far⁴⁹. A single dose was shown to be uniformly protective against lethal TAFV challenge with the vaccinated macaques not developing any signs of disease or viremia⁵⁰. As stated above, this vaccine had been shown to provide limited cross-protection against BDBV infection⁴⁵. The VSVΔG platform vaccines had previously demonstrated cross-protection against TAFV infection when a 1:1:1 blend of 3 VSV vaccines (VSV-EBOV, VSV-SUDV and VSV-MARV) was administered 4 weeks before lethal challenge. Uniform protection was achieved in the VSV blend group compared to 40% in the control group⁴⁰. TAFV GP antibodies have also been demonstrated with Ad26 and Ad35 vaccination blending vectors expressing multiple filovirus GPs, however, no TAFV challenge was performed⁵¹. Lastly, MVA-BN-filo vector expresses the TAFV NP in combination with the SUDV GP, EBOV GP, and MARV GP but has never been evaluated against lethal TAFV challenge⁵². The cross-protective potential of this vaccine vector in combination with Ad26 vectors is discussed below.

MARV vaccines

While MARV was the first filovirus to be discovered in 1967, it has not caused as many outbreaks of human disease compared to EBOV⁵³. The two major outbreaks occurred in the Democratic Republic of Congo from 1997–2000 (154 cases, 128 fatalities) and Angola 2004–2005 (252 cases, 227 fatalities), yet MARV has not been as well-studied as EBOV⁵³. This is also reflected in the progress made in countermeasure development. To date, there are 6 MARV-specific vaccines in preclinical and clinical development—one subunit vaccine, 3 vaccines based on the VSV platform, and 2 on the Ad platform. The subunit vaccine is composed of MARV GP and CoVaccine HT adjuvant and has demonstrated uniform protection in macaques after 3 doses¹⁹. This vaccine does not confer a rapid onset of protection, but the protective responses may be more durable. The VSV-N4CT1-MARV from Auro vaccines, a genetically altered VSV platform including the VSV glycoprotein, expresses the MARV-Angola GP as an additional open reading frame leading to uniform protection when a single high dose is administered prior to lethal MARV challenge. If this vaccine is administered closer to challenge (5 or

3 days prior), efficacy drops to 80% or 20% survival, respectively⁵⁴. This is in contrast to VSV-MARV (expressing the MARV-Angola GP on the VSVΔG platform) for which vaccination with a single high dose even 3 days prior to infection leads to 75% survival in lethally MARV challenged macaques¹⁵. This vaccine is highly immunogenic and protects by eliciting a humoral as well as cellular responses⁵⁵. Recently, this vaccine demonstrated uniform protection at a low dose within 7 days⁵⁶, and is currently in clinical development by Public Health Vaccines with clinical trial data pending (NCT06265012)²⁶. The last VSV-based MARV vaccine is the original version of VSV-MARV expressing the MARV-Musoke GP in the VSVΔG platform which demonstrated uniform protection in macaques within 28 days¹⁰ and durability of the uniformly protective response up to 407 days⁵⁷. This vaccine is in clinical development by the International AIDS Vaccine Initiative²⁶.

The 2 non-replicating viral vector vaccines are based on ChAd/cAd, namely ChAdOx-MARV (Oxford University) and cAd3-MARV (Sabin), expressing the MARV-Angola GP. There is no published data for the ChAdOx-MARV vaccine, but the cAd3-MARV has been shown to offer rapid (7 days before challenge) and durable (6 months before challenge) uniform protection in macaques with a single high dose¹⁸. After 12 months, the survival rate in single dose vaccinated macaques drops to 75%¹⁸. The cAd3-MARV is the only vaccine with available human clinical trial data. In a phase 1 trial in the USA the vaccine was shown to be immunogenic with an acceptable safety profile at doses of 1×10^{10} and 1×10^{11} PU (NCT03475056)⁵⁸. Of the 40 participants 95% developed MARV GP-specific IgG 4 weeks after vaccination and the response remained measurable in 70% of the participants at 48 weeks⁵⁸. A phase 2 clinical trial is currently ongoing in Uganda (NCT05817422).

Multivalent filovirus vaccines

Since several human-pathogenic filoviruses overlap in their endemic areas of West, Central and East Africa, there is a push towards the development of pan-filovirus vaccines. Currently, there are several platforms in development for multi-valent approaches.

Bivalent subunit vaccines combining the insect cell-expressed GPs of EBOV and SUDV or EBOV and MARV with the adjuvant CoVaccine HT have been shown to uniformly protect macaques from lethal EBOV, SUDV, and MARV challenge after 3 doses¹⁹. In addition, the adjuvanted SUDV GP and MARV GP combination has uniformly protected SUDV- or MARV-challenged macaques after 3 vaccine doses⁴¹. Both formulations of these bivalent vaccines do not confer a rapid onset of protection, however, they are suitable for population-based vaccination particularly in the thermostable formulation. As adjuvanted subunit vaccines, adverse events are expected to be limited, however, the required 3 doses for protective efficacy presents a challenge for roll-out in rural filovirus endemic areas.

As stated above, while presumably immunogenic in a phase I clinical trial, a ChAdOx-biEBOV expressing both the EBOV GP and SUDV GP was not protective against lethal SUDV infection in macaques and development has ceased³⁶. Based on the Ad26 platform, a trivalent filovirus vaccine consisting of Ad26-filo (a 1:1:1 blend of Ad26-EBOV, Ad26-SUDV and Ad26-MARV) and MVA-BN-filo has been shown to protect as a prime/boost strategy against lethal SUDV and MARV challenge in NHPs⁵². Notably, when MVA-BN-filo was administered first and Ad26-filo used as a boost, the protective efficacy decreased⁵². The safety and immunogenicity of doses as high as 9×10^{10} PU of Ad26-filo and 5×10^8 infectious units of MVA-BN-filo administered 14 or 56 days apart was evaluated in a phase 1 clinical trial (NCT02860650). The study reports no serious adverse events after vaccination and 21 days after the boost vaccination all study participants had measurable binding antibody serum titers to the 3 filovirus GP antigens²⁰. Neutralization activity was robust for EBOV, but suboptimal for SUDV and MARV²⁰. This vaccine has not yet been moved further along in clinical development.

The VSV platform has also been explored for mediating pan-filovirus immunity. In 2009, single-dose vaccination with a blend of VSV-EBOV,

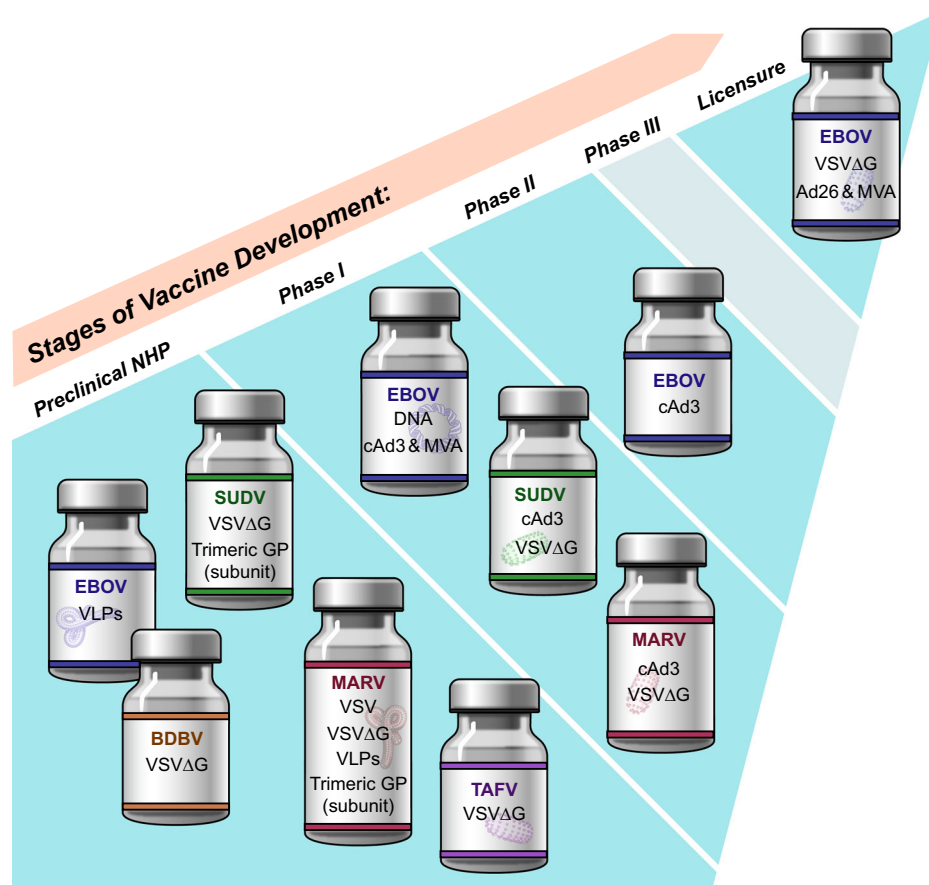


Fig. 1 | Development stages of filovirus vaccines in the USA. The cartoon indicates the development stages of different filovirus vaccines. Credit: NIAID.

VSV-SUDV and VSV-MARV provided uniform protection to macaques challenged with EBOV, SUDV, MARV or the heterologous TAFV⁴⁰. However, no further studies have followed up on this concept using the VSVΔG vector. In contrast, Auro vaccines developed their VSV vector (VSV-N4CT1 including the VSV glycoprotein) into a trivalent formulation by blending vectors expressing EBOV GP, SUDV GP and MARV GP in a 1:1:1 ratio. A single dose protected all macaques from lethal challenge with EBOV, SUDV, or MARV⁵⁹. Because filoviruses share their endemic areas with Lassa virus (LASV), the next study examining protection included a VSV vector expressing the LASV GP in the previously described trivalent blend. Administration of 2 doses (56 days apart) of this tetravalent blend (1:1:1:1 ratio) was uniformly protective against lethal challenge with EBOV, SUDV, MARV, and LASV in macaques⁶⁰. Most recently, a study investigating the rapid onset of protection using a pan-filovirus vaccine (1:1:1:1 blend of VSV-N4CT1-EBOV, VSV-N4CT1-SUDV, VSV-N4CT1-MARV, and VSV-N4ΔG-BDBV) reported uniform protection within 7 days of vaccination for macaques challenged with SUDV, BDBV and MARV; however, EBOV challenge resulted in only 80% survival⁶¹.

Future challenges

Filovirus vaccine development has generated licensed products for EVD but products for other human pathogenic filoviruses are not even close to the finish line. Efforts are being intensified to bring the most promising products, particularly for SUDV and MARV, from preclinical to clinical development and to licensure to cover potential reemergence in the African endemicity zones. These efforts include several new vaccine approaches based on updated platforms and new technologies which have yet to demonstrate preclinical efficacy in NHPs. In addition, mediation strategies need also be implemented for truly emerging, novel human-pathogenic filoviruses. As the

development of these countermeasures is dependent on the continued infectious disease and public health interest and funding, efforts communicating these needs and bringing them to the stakeholders' attention is critical.

Vaccine production for clinical use and vaccine storage for emergency release during outbreaks are challenging issues. The most effective way would likely be production in a developed country to use existing knowledge and facilities and storage in endemic regions for immediate and hopefully less complicated release in case of outbreaks. Currently largely unanswered questions relate to shelf-life time, storage temperature, and cold chain requirements of the different products. Continuation of these efforts requires further studies and appropriate funding.

Cross-protective vaccines would be helpful to cover the variation among human pathogenic filoviruses. However, single cross-protective vaccines would be preferred but are currently not available. Currently, cross-protective approaches include heterologous prime-boost and blending of vaccine candidates. Preclinical cross-protection studies may be difficult to interpret as protective efficacy is easier demonstrated in rodent than macaque models⁶². Taking this a step further, an extrapolation of cross-protective responses from macaques to humans is complicated as animal models use artificial exposure routes including intramuscular and intra-peritoneal while natural filovirus infection is believed to occur mucosally¹. Recent studies demonstrating disease in macaques after mucosal exposure are aiding in addressing this knowledge gap^{63,64}.

Except for MVA-BN-filo, currently all filovirus vaccines, approved or in development, use the GP as the viral immunogen. Other viral proteins have been evaluated as immunogens especially for EBOV but with limited success⁶⁵. Data from existing studies mapping immune responses to epitopes and other insightful details may inform the design of antigens especially those suitable to elicit cross-protective immune responses.

Finally, a rapid response strategy for vaccine development would be needed to address truly novel emerging filoviruses. Ideally, this would be based on existing successful vaccine platforms. To realize this effort, uncomplicated and immediate sharing of infectious material or virus isolates would be a prerequisite but is currently often not achievable.

Conclusion

Since the 2013–2016 West African EVD epidemic the filovirus field has come a long way with approved diagnostic tests, vaccines, and treatments against EBOV. Efforts are ongoing to reach a similar status with approved countermeasure for SUDV and MARV. There are several approaches in clinical development with promising initial results particularly on the vaccine front and the most recent outbreaks have revived interest and enabled funding¹². These efforts should be expanded to include BDBV and TAFV as they remain the only known human-pathogenic filoviruses without a countermeasure landscape in considerable stages of development (Fig. 1). Along the same lines, studies should be intensified to define the pathogenic potential of novel filoviruses.

While the field is moving into the right direction, one of the biggest challenges remaining is the timeliness of the public health response after outbreaks are declared. Agreements are needed between countermeasure stakeholders, regulatory authorities, and governments to establish a framework particularly in the endemic areas in Africa to enable a faster public health response and deployment of vaccines and treatments to control disease outbreaks. A prerequisite for success is the involvement of African countries and their public health infrastructure.

Finally, lessons learned from filovirus vaccine development may guide responses to other infectious disease outbreaks, especially those caused by zoonotic pathogens. Response strategies, however, must be tailored to the individual pathogen/disease and there is no universal protocol that fits all. For instance, outbreaks caused by the likely bat-borne filoviruses are largely driven by human-to-human transmission, whereas outbreaks driven by the rodent-borne orthohantaviruses and mammarenaviruses are largely driven by rodent-to-human and only limited human-to-human transmission. Yet, implementations of vaccines can follow an analogous pathway for which EBOV can serve as a response paradigm. Therefore, we should ensure that preclinical development of vaccines for prioritized pathogens is accomplished including clinical grade material production, product storage, and arrangements for instantaneous, uncomplicated deployment. This will allow emergency clinical assessment during outbreaks similar to what was done in West African during the EVD epidemic.

Received: 16 June 2024; Accepted: 25 September 2024;
Published online: 11 October 2024

References

- Jacob, S. T. et al. Ebola virus disease. *Nat. Rev. Dis. Prim.* **6**, 13 (2020).
- Towner, J. S. et al. Isolation of genetically diverse Marburg viruses from Egyptian fruit bats. *PLoS Pathog.* **5**, e1000536 (2009).
- Schindell, B. G., Webb, A. L. & Kindrachuk, J. Persistence and sexual transmission of filoviruses. *Viruses* **10**, <https://doi.org/10.3390/v10120683> (2018).
- Durant, O. & Marzi, A. Ebola virus disease sequelae and viral persistence in animal models: Implications for the future. *PLoS Pathog.* **20**, e1012065 (2024).
- Anderson, E. M. & Collier, B. A. Translational success of fundamental virology: a VSV-vectored Ebola vaccine. *J. Virol.* e0162723. <https://doi.org/10.1128/jvi.01627-23> (2024).
- Tomori, O. & Kolawole, M. O. Ebola virus disease: current vaccine solutions. *Curr. Opin. Immunol.* **71**, 27–33 (2021).
- Lee, A. Ansumivab: first approval. *Drugs* **81**, 595–598 (2021).
- Rayaprolu, V. et al. Structure of the Inmazeb cocktail and resistance to Ebola virus escape. *Cell Host Microbe* **31**, 260–272.e267 (2023).
- Mulangu, S., Mbala-Kingebeni, P. & Mbaya, O. T. Antibody use during an outbreak of Ebola virus disease in the Democratic Republic of Congo, 2020. *N. Engl. J. Med.* **386**, 1188–1191 (2022).
- Jones, S. M. et al. Live attenuated recombinant vaccine protects nonhuman primates against Ebola and Marburg viruses. *Nat. Med.* **11**, 786–790 (2005).
- Izudi, J. & Bajunirwe, F. Case fatality rate for Ebola disease, 1976–2022: a meta-analysis of global data. *J. Infect. Public Health* **17**, 25–34 (2024).
- Parish, L. A., Stavale, E. J., Houchens, C. R. & Wolfe, D. N. Developing vaccines to improve preparedness for filovirus outbreaks: the perspective of the USA biomedical advanced research and development authority (BARDA). *Vaccines* **11**, <https://doi.org/10.3390/vaccines11061120> (2023).
- Marzi, A. et al. EBOLA VACCINE. VSV-EBOV rapidly protects macaques against infection with the 2014/15 Ebola virus outbreak strain. *Science* **349**, 739–742 (2015).
- Henao-Restrepo, A. M. et al. Efficacy and effectiveness of an rVSV-vectored vaccine in preventing Ebola virus disease: final results from the Guinea ring vaccination, open-label, cluster-randomised trial (Ebola Ca Suffit). *Lancet* **389**, 505–518 (2017).
- Marzi, A. et al. Single dose of a VSV-based vaccine rapidly protects macaques from marburg virus disease. *Front. Immunol.* **12**, 774026 (2021).
- Marzi, A. & Feldmann, H. Ebola virus vaccines: an overview of current approaches. *Expert Rev. Vaccines* **13**, 521–531 (2014).
- Woolsey, C. et al. A recombinant vesicular stomatitis virus-based vaccine provides postexposure protection against bundibugyo ebolavirus infection. *J. Infect. Dis.* **228**, S712–S720 (2023).
- Hunegnaw, R. et al. A single-shot ChAd3-MARV vaccine confers rapid and durable protection against Marburg virus in nonhuman primates. *Sci. Transl. Med.* **14**, eabq6364 (2022).
- Lehrer, A. T. et al. Recombinant protein filovirus vaccines protect cynomolgus macaques from Ebola, Sudan, and Marburg Viruses. *Front. Immunol.* **12**, 703986 (2021).
- Bockstal, V. et al. First-in-human study to evaluate safety, tolerability, and immunogenicity of heterologous regimens using the multivalent filovirus vaccines Ad26.Filo and MVA-BN-Filo administered in different sequences and schedules: a randomized, controlled study. *PLoS ONE* **17**, e0274906 (2022).
- de La Vega, M. A. et al. An update on nonhuman primate usage for drug and vaccine evaluation against filoviruses. *Expert Opin. Drug Discov.* <https://doi.org/10.1080/17460441.2024.2386100> (2024).
- van Tol, S. et al. A bivalent Adenovirus-Vectored Vaccine induces a robust humoral response, but does not protect cynomolgus macaques against a lethal challenge with Sudan virus. *J. Infect. Dis.* <https://doi.org/10.1093/infdis/jiae056> (2024).
- Kemenesi, G. et al. Isolation of infectious Lloviu virus from Schreiber's bats in Hungary. *Nat. Commun.* **13**, 1706 (2022).
- Bodmer, B. S. et al. In vivo characterization of the novel ebolavirus Bombali virus suggests a low pathogenic potential for humans. *Emerg. Microbes Infect.* **12**, 2164216 (2023).
- Fletcher, P. et al. Pathogenicity of Lloviu and Bombali Viruses in Type I interferon receptor knockout mice. *J. Infect. Dis.* **228**, S548–S553 (2023).
- Cross, R. W. et al. An introduction to the Marburg virus vaccine consortium, MARVAC. *PLoS Pathog.* **18**, e1010805 (2022).
- Branswell, H. As foreign powers approve Ebola vaccines, U.S. drug makers lag in development pipeline. <https://www.statnews.com/2017/12/08/ebola-vaccine-development/> (2017).
- WHO. Ebola virus disease: Vaccines, <https://www.who.int/news-room/questions-and-answers/item/ebola-vaccines> (2020).
- Merck. <https://www.merck.com/news/u-s-fda-approves-mercks-ervebo-ebola-zaire-vaccine-live-for-use-in-children-12-months-of-age-and-older/>, <https://www.merck.com/news/european-commission-expands-mercks-ervebo-ebola-zaire-vaccine-rvsv%>

- CE%B4gzebov-gp-live-indication-to-include-children-1-year-of-age-and-older/ (2023).
30. WHO. *Four countries in the African region license vaccine in milestone for Ebola prevention*, <https://www.who.int/news/item/14-02-2020-four-countries-in-the-african-region-license-vaccine-in-milestone-for-ebola-prevention> (2020).
31. Kallay, R. et al. Use of Ebola vaccines - worldwide, 2021–2023. *MMWR Morb. Mortal. Wkly Rep.* **73**, 360–364 (2024).
32. Coulborn, R. M. et al. Case fatality risk among individuals vaccinated with rVSVDeltaG-ZEBOV-GP: a retrospective cohort analysis of patients with confirmed Ebola virus disease in the Democratic Republic of the Congo. *Lancet Infect. Dis.* [https://doi.org/10.1016/S1473-3099\(23\)00819-8](https://doi.org/10.1016/S1473-3099(23)00819-8) (2024).
33. CDC. *Ebola (Ebola virus disease) - Outbreaks*, <https://www.cdc.gov/vhf/ebola/history/chronology.html> (2023).
34. Nakkazi, E. A trial for Ebola Sudan virus in Uganda. *Lancet Infect. Dis.* **23**, 158 (2023).
35. Bwire, G. et al. Sudan Ebola virus (SUDV) outbreak in Uganda, 2022: lessons learnt and future priorities for sub-Saharan Africa. *BMC Med.* **21**, 144 (2023).
36. van Tol, S. et al. A bivalent Adenovirus-Vectorized Vaccine induces a measurable humoral response, but does not protect cynomolgus macaques against a lethal challenge with Sudan virus. *bioRxiv.org* <https://doi.org/10.1101/2023.10.20.563337> (2023).
37. Honko, A. N. et al. A single-shot ChAd3 vaccine provides protection from intramuscular and aerosol sudan virus exposure. *bioRxiv* <https://doi.org/10.1101/2024.02.07.579118> (2024).
38. Mwesigwa, B. et al. Safety, tolerability, and immunogenicity of the Ebola Sudan chimpanzee adenovirus vector vaccine (cAd3-EBO S) in healthy Ugandan adults: a phase 1, open-label, dose-escalation clinical trial. *Lancet Infect. Dis.* **23**, 1408–1417 (2023).
39. Marzi, A. et al. Species-specific immunogenicity and protective efficacy of a vesicular stomatitis virus-based Sudan virus vaccine: a challenge study in macaques. *Lancet Microbe* [https://doi.org/10.1016/S2666-5247\(23\)00001-0](https://doi.org/10.1016/S2666-5247(23)00001-0) (2023).
40. Geisbert, T. W. et al. Single-injection vaccine protects nonhuman primates against infection with marburg virus and three species of ebola virus. *J. Virol.* **83**, 7296–7304 (2009).
41. To, A. et al. Thermostable bivalent filovirus vaccine protects against severe and lethal Sudan ebolavirus and marburgvirus infection. *Vaccine* **42**, 598–607 (2024).
42. Towner, J. S. et al. Newly discovered ebola virus associated with hemorrhagic fever outbreak in Uganda. *PLoS Pathog.* **4**, e1000212 (2008).
43. MacNeil, A. et al. Proportion of deaths and clinical features in Bundibugyo Ebola virus infection, Uganda. *Emerg. Infect. Dis.* **16**, 1969–1972 (2010).
44. Kratz, T. et al. Ebola virus disease outbreak in Isiro, Democratic Republic of the Congo, 2012: signs and symptoms, management and outcomes. *PLoS ONE* **10**, e0129333 (2015).
45. Falzarano, D. et al. Single immunization with a monovalent vesicular stomatitis virus-based vaccine protects nonhuman primates against heterologous challenge with Bundibugyo ebolavirus. *J. Infect. Dis.* **204**, S1082–S1089 (2011).
46. Mire, C. E. et al. Vesicular stomatitis virus-based vaccines protect nonhuman primates against Bundibugyo ebolavirus. *PLoS Neglect. Trop. Dis.* **7**, e2600 (2013).
47. Formenty, P. et al. Human infection due to Ebola virus, subtype Cote d'Ivoire: clinical and biologic presentation. *J. Infect. Dis.* **179**, S48–S53 (1999).
48. CDC. *Marburg Disease Outbreaks*, <https://www.cdc.gov/vhf/marburg/outbreaks/chronology.html> (2023).
49. Marzi, A. et al. Vesicular stomatitis virus-based Ebola vaccines with improved cross-protective efficacy. *J. Infect. Dis.* **204**, S1066–S1074 (2011).
50. Fletcher, P. et al. Single-dose VSV-based vaccine protects cynomolgus macaques from disease after Tai Forest virus infection. *Emerg. Microbes Infect.* **12**, 2239950 (2023).
51. Callendret, B. et al. A prophylactic multivalent vaccine against different filovirus species is immunogenic and provides protection from lethal infections with Ebolavirus and Marburgvirus species in non-human primates. *PLoS ONE* **13**, e0192312 (2018).
52. Tiemessen, M. M. et al. Protection against Marburg virus and Sudan virus in NHP by an adenovector-based trivalent vaccine regimen is correlated to humoral immune response levels. *Vaccines* **10**, <https://doi.org/10.3390/vaccines10081263> (2022).
53. Slenczka, W. & Klenk, H. D. Forty years of Marburg virus. *J. Infect. Dis.* **196**, S131–S135 (2007).
54. Woolsey, C. et al. A highly attenuated Vesiculovax vaccine rapidly protects nonhuman primates against lethal Marburg virus challenge. *PLoS Neglect. Trop. Dis.* **16**, e0010433 (2022).
55. Marzi, A. et al. Protection against Marburg virus using a recombinant VSV-vaccine depends on T and B cell activation. *Front. Immunol.* **9**, 3071 (2018).
56. O'Donnell, K. L. et al. Rapid protection of nonhuman primates against Marburg virus disease using a single low-dose VSV-based vaccine. *EBioMedicine* **89**, 104463 (2023).
57. Mire, C. E. et al. Durability of a vesicular stomatitis virus-based Marburg virus vaccine in nonhuman primates. *PLoS ONE* **9**, e94355 (2014).
58. Hamer, M. J. et al. Safety, tolerability, and immunogenicity of the chimpanzee adenovirus type 3-vectorized Marburg virus (cAd3-Marburg) vaccine in healthy adults in the USA: a first-in-human, phase 1, open-label, dose-escalation trial. *Lancet* **401**, 294–302 (2023).
59. Matassov, D. et al. Single-dose trivalent VesiculoVax vaccine protects macaques from lethal ebolavirus and marburgvirus challenge. *J. Virol.* **92**, <https://doi.org/10.1128/JVI.01190-17> (2018).
60. Cross, R. W. et al. Quadrivalent VesiculoVax vaccine protects nonhuman primates from viral-induced hemorrhagic fever and death. *J. Clin. Invest.* **130**, 539–551 (2020).
61. Woolsey, C. et al. A highly attenuated panfilovirus VesiculoVax vaccine rapidly protects nonhuman primates against marburg virus and 3 species of Ebola virus. *J. Infect. Dis.* **228**, S660–S670 (2023).
62. Marzi, A., Feldmann, H., Geisbert, T. W. & Falzarano, D. Vesicular stomatitis virus-based vaccines for prophylaxis and treatment of filovirus infections. *J. Bioterrorism Biodefense* **S1**, <https://doi.org/10.4172/2157-2526.S1-004> (2011).
63. Mire, C. E. et al. Oral and conjunctival exposure of nonhuman primates to low doses of Ebola Makona Virus. *J. Infect. Dis.* **214**, S263–S267 (2016).
64. Prasad, A. N. et al. Natural history of nonhuman primates after oral exposure to Ebola Virus Variant Makona. *J. Infect. Dis.* **228**, S571–S581 (2023).
65. Reynolds, P. & Marzi, A. Ebola and Marburg virus vaccines. *Virus Genes* **53**, 501–515 (2017).

Acknowledgements

The authors thank Rose Gottschalk-Perry (NIAID) for assistance with figure production using Adobe Illustrator and colleagues in the field for fruitful discussion and suggestions. A.M. and H.F. are supported by the Intramural Research Program of the NIAID, NIH.

Author contributions

A.M. and H.F. equally developed the concept of the review article, generated the drafts for the display items, and contributed to writing of the manuscript. A.M. and H.F. equally secured funding. All authors read and approved the final manuscript.

Funding

Open access funding provided by the National Institutes of Health.

Competing interests

A.M. declares no competing interests. H.F. claims intellectual property for VSV-based filovirus vaccines.

Additional information

Correspondence and requests for materials should be addressed to Andrea Marzi or Heinz Feldmann.

Reprints and permissions information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

This is a U.S. Government work and not under copyright protection in the US; foreign copyright protection may apply 2024